

ApoTox-Glo™ Triplex Assay

Assay for live, dead and apoptotic cells in the same well

- [Assay Principle](#)
- [Protocol Overview](#)
- [Necrosis or Apoptosis](#)
- [Sensitivity](#)
- [Flexibility](#)
- [Ordering Information](#)
- [References](#)

- ✓ Sensitive, non-lytic, fluorescent assays quantitate live and dead cells
- ✓ Sensitive luminescent assay for caspase 3/7 activation.
- ✓ Simple two-step add-mix-measure protocols
- ✓ Adaptable to 96-, 384- & 1,536-well plate assays



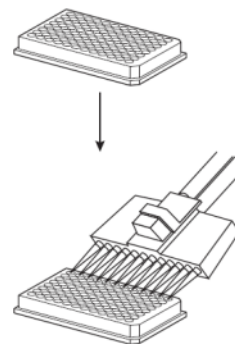
Protocol Overview

- [Principle](#)
- [Protocol](#)
- [Sensitivity](#)
- [Flexibility](#)
- [Apoptosis?](#)
- [Ordering](#)
- [References](#)



Go to the
ApoTox-Glo™
Triplex Assay
Technical
Manual

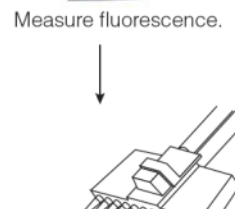
Measure
AFC fluorescence
380-400nm_{EX}/505nm_{EM}
R110 fluorescence
485nm_{EX}/520nm_{EM}



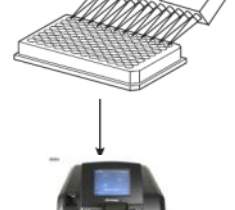
Treat cells grown in black or white plates with compound of interest (typically, 0-24 hours)



Add Cytotoxicity/Viability Reagent.
Mix & incubate 30 minutes

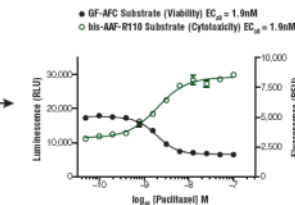


Add Caspase-Glo® 3/7 Reagent.
Mix & incubate 30 minutes

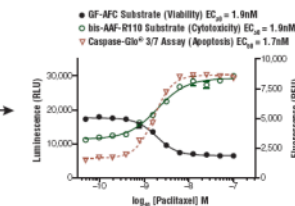


Measure
Luminescence

Measure luminescence.

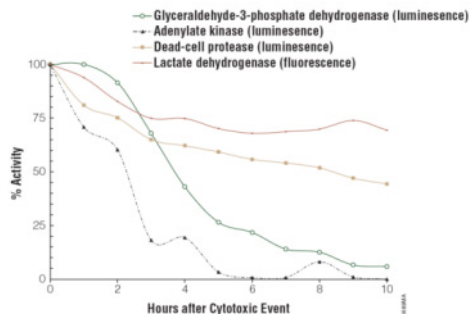


Ratiometric
Live/Dead Cells



Live, Dead and
Apoptotic Cells

Timing of assays to measure a released cellular enzyme is critical. You must consider the half-life of the enzyme outside the cell.



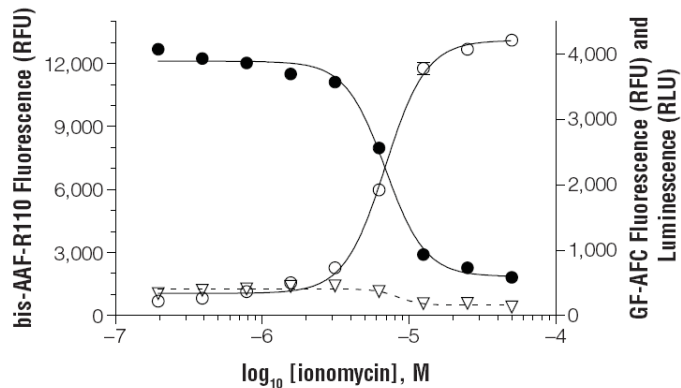
Half-life of enzymatic
markers for cytotoxicity



Necrosis or Apoptosis defined

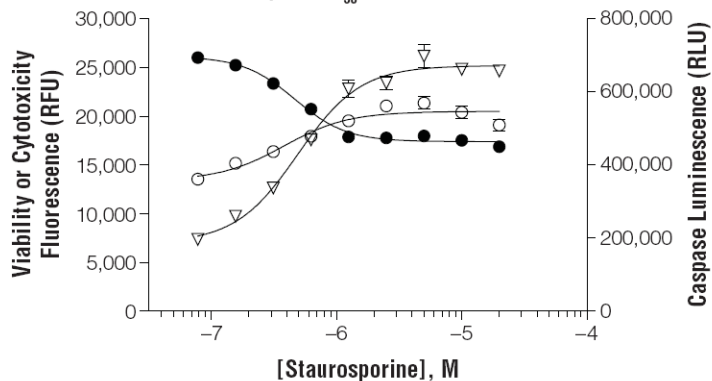
- [Principle](#)
- [Protocol](#)
- [Sensitivity](#)
- [Flexibility](#)
- Apoptosis?
- [Ordering](#)
- [References](#)

- GF-AFC (Viability) $EC_{50} = 6.89\mu M$
- bis-AAF-R110 (Cytotoxicity) $EC_{50} = 6.87\mu M$
- ▽ Caspase-Glo® 3/7 Assay (Apoptosis) $EC_{50} = N.D.$



Necrotic Cell Death
Little or no caspase-3/7 activation

- Viability $EC_{50} = 463nM$
- Cytotoxicity $EC_{50} = 380nM$
- ▽ Caspase $EC_{50} = 491nM$



Apoptotic Cell Death
Caspase-3/7 activation

Jurkat cells treated for 6 hours with either ionomycin or staurosporine



Sensitivity

[Principle](#)

[Protocol](#)

[Sensitivity](#)

[Flexibility](#)

[Apoptosis?](#)

[Ordering](#)

[References](#)

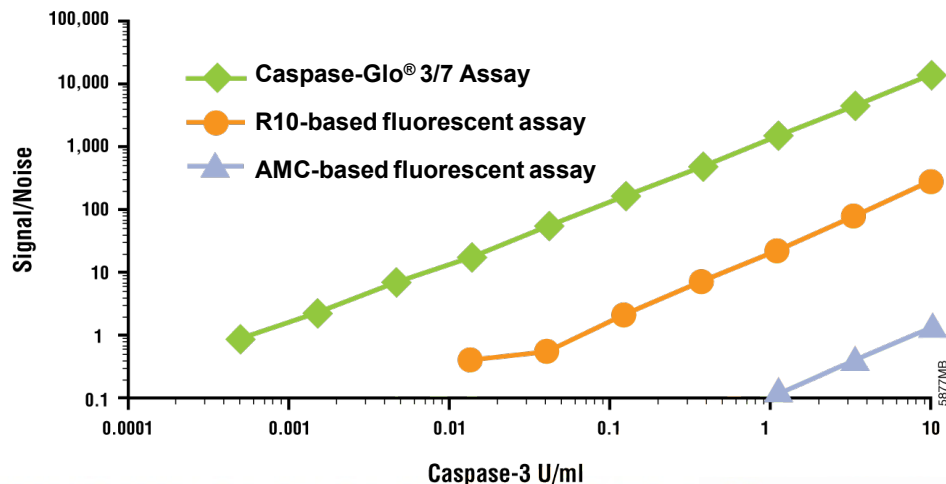
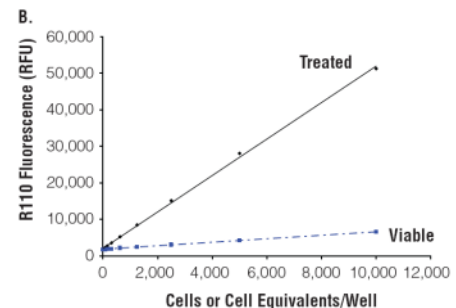
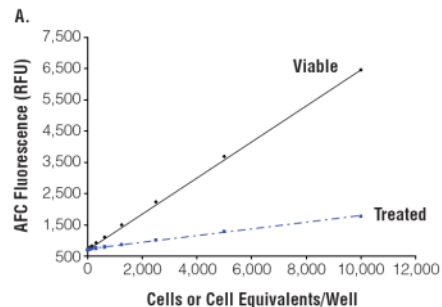
A pool of Jurkat cells was divided into two fractions. One fraction was treated to simulate cytotoxicity, whereas the other was left untreated.

Detect caspase-3/7 activation earlier and with fewer cells.

The Caspase-Glo® 3/7 Assay can detect caspase 3/7 activation at lower levels than fluorescent methods.

Viability Assay
Live Cell Detection Limits
96-well: ~400 cells
384-well: ~50 cells

Cytotoxicity Assay
Dead Cell Detection
Limits 2-5% dead cells
per well



Flexibility for High-Throughput Assays

The ApoTox-Glo Assay combines the MultiTox-Fluor and Caspase-Glo® 3/7 Assays giving tight CVs and excellent Z' values.

[Principle](#)

[Protocol](#)

[Sensitivity](#)

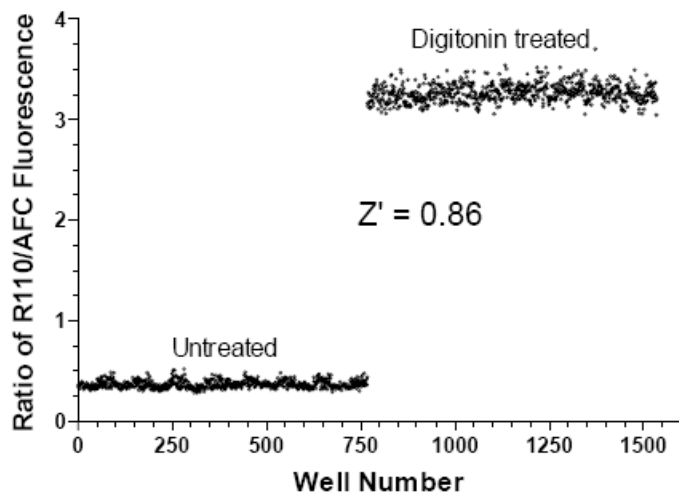
[Flexibility](#)

[Apoptosis?](#)

[Ordering](#)

[References](#)

MultiTox-Fluor 1536 Z'-Factor



Full plates of Jurkat cells (2,500/well 1,536-well plates) were prepared. Half of each plate was left untreated and the other half got digitonin. The cells were assayed simultaneously for dead cells (CytoTox-Fluor™ Assay; AAF-Rhodamine 110 substrate) and live cells (CellTiter-Fluor™ Assay; GF-AFC substrate).

Source: [Niles, A., Worzella, T. and Busch, M. \(2008\) Automation of multiplexed cell-based viability and cytotoxicity assays. Presented at the Society for Biomolecular Sciences \(SBS\).](#)

Caspase-Glo® 3/7 Assay

Format	Assay Volume	Cells/Well	%CV ¹	Z' Factor ²
1536	8µl	2000	5.27	0.77
	5µl	1250	5.07	0.79
	2µl	500	7.78	0.71
	1µl	250	6.12	0.62
	0.5µl	125	8.53	n.t.

¹ The %CV data was obtained by plating a serial dilution of Jurkat or d293 cells and inducing apoptosis with either anti-FAS antibody or Triton®-X 100. The Caspase-Glo 3/7 Reagent was added and light units recorded. The %CV values listed here were calculated from the maximum cell number used in each titration series.

² The Caspase-Glo 3/7 Z'-factor assays were performed by plating Jurkat cells and treating one-half of the plate with an anti-FAS antibody for four hours, with the remaining half receiving no treatment. The Caspase-Glo 3/7 Reagent was then added and light units were recorded.

Source: [Worzella, T., et al. \(2006\) Automating Promega CellTiter-Glo® Luminescent Cell Viability and Caspase-Glo® 3/7 Assays in Low-Volume 384 and 1536-Well Formats. Society for Biomolecular Sciences \(SBS\) meeting poster.](#)



Ordering Information

- [Principle](#)
- [Protocol](#)
- [Sensitivity](#)
- [Flexibility](#)
- [Apoptosis?](#)
- [Ordering](#)
- [References](#)

Product	Size	Catalog Number
ApoTox-Glo™ Triplex Assay ^(a,b,c)	10ml	PRG6320
	5 x 10ml	PRG6321

For Laboratory Use. G6320 contains sufficient reagents for 100 assays at 100µl/assay in a 96-well plate format or 400 assays at 25µl/assay in a 384-well format. G6321 contains sufficient reagents for 500 assays at 100µl/assay in a 96-well plate format or 2,000 assays at 25µl/assay in a 384-well format.



For customer service, call 1-800-766-7000.
To fax an order, use 1-800-926-1166.
To order online: www.fishersci.com

©2009 Thermo Fisher Scientific

Questions?

e-mail: Technical Services
techserv@promega.com

800-356-9526, Option 4
Available 8am-7pm Eastern
Monday-Friday



Links to more information

[Principle](#)
[Protocol](#)
[Sensitivity](#)
[Flexibility](#)
[Apoptosis?](#)
[Ordering](#)
[References](#)

ApoTox-Glo™ Assay



[Shultz, S., et al. \(2009\) Automated triplex assay to assess cell viability, cytotoxicity and apoptosis. Presented at Lab Automation 2009 Conference.](#)

MultiTox-Fluor and/or Caspase-Glo® 3/7 Assay



[Niles, A., et al. \(2006\) Multiplexed viability, cytotoxicity and apoptosis assays for cell-based screening. *Cell Notes* **16**, 12-15.](#)



[Niles, A., et al. \(2008\) Using protease biomarkers to measure viability and cytotoxicity. *Cell Notes* **19**, 16-20.](#)



[Zakowicz, H., et al. \(2008\) Measuring cell health and viability sequentially by same-well multiplexing using the GloMax®-Multi Detection System. *Promega Notes* **99**, 25-28.](#)



[Worzella, T., Busch, M. and Niles, A. \(2008\) High-throughput automation of multiplexed cell-based assays for viability and cytotoxicity. *Cell Notes* **20**, 26-29.](#)



[Niles, A.L., et al. \(2006\) MultiTox-Fluor Multiplex Cytotoxicity Technology. *Cell Notes* **15**, 11-15.](#)



[Niles, A., et al. \(2007\) Measure relative numbers of live and dead cells and normalize assay data to cell number. *Cell Notes* **18**, 15-20.](#)



[He, J.-Q., Ma, J. and Anson, B. \(2009\) Use of pluripotent stem cell-derived cardiomyocytes to understand mechanisms of cardiotoxic compounds. *Cell Notes* **23**, 10-13.](#)



[O'Brien, M., Moravec, R. and Riss, T. \(2003\) Caspase-Glo 3/7 Assay: Use fewer cells and spend less time with this homogeneous assay. *Cell Notes* **6**, 13-15.](#)

[MultiTox-Fluor HighWire Press®](#)

[MultiTox-Fluor from Nature.com](#)

[MultiTox-Fluor PubChem BioAssays](#)

[Caspase-Glo 3/7 HighWire Press®](#)

[Caspase-Glo 3/7 from Nature.com](#)

[Caspase-Glo 3/7 PubChem BioAssays](#)

